

SEQUENCE LISTING

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<120> Novel Human Cancer Antigen NY ESO-1/CAG-3 and Gene Encoding Same

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<140> PCT/US98/19609
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<141> 1998-09-21

<150> US60/061,428

<151> 1997-10-08

<160> 106

<170> PatentIn Ver. 2.0

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Gly Ala Ala Arg Ala Ser Gly Pro Gly Gly Gly Ala Pro Arg Gly Pro
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His Gly Gly Ala Ala Ser Gly Leu Asn Gly Cys Cys Arg Cys Gly Ala
65 70 75 80

Arg Gly Pro Glu Ser Arg Leu Leu Glu Phe Tyr Leu Ala Met Pro Phe 85 90 95

Ala Thr Pro Met Glu Ala Glu Leu Ala Arg Arg Ser Leu Ala Gln Asp 100 105 110

Ala Pro Pro Leu Pro Val Pro Gly Val Leu Leu Lys Glu Phe Thr Val 115 120 125 Ser Gly Asn Ile Leu Thr Ile Arg Leu Thr Ala Ala Asp His Arg Gln 130 135 140

Leu Gln Leu Ser Ile Ser Ser Cys Leu Gln Gln Leu Ser Leu Leu Met 145 150 155 160

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      acids, Xaa2 is Ala, Thr, Val, Leu or Arg, Xaa3 is
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                                   25
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Reverse Transcriptase-PCR

Total RNA was extracted from tumor cell lines as described above. Five hundred nanograms of total RNA was used for conversion of RNA to cDNA by avian myeloblastosis virus (AMV) reverse transcriptase. cDNA was then amplified by PCR using the ESO-P2 (5'GCGGCTTCAGGGCTGAATGGATG) (SEQ. ID No. 105) and ESO-P5 (5'-AAGCCGTCCTCCTCCAGCGACA) (SEQ. ID No. 106) primers and One-Step RT-PCT system (Life Technologies). PCR products were amplified under denaturation conditions at 94°C for 30 s, annealing at 55° for 30 s, extension at 72° for 3 min for 40 cycles, and final elongation at 72° for 10 min. PCR products were analyzed on a 3% agarose gel.

Cytotoxic lysis assays

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Cytolytic assay was done as previously described (Kawakami, Y et al., 1994, Proc. Natl. Acad. Sci. USA 91:6458-62). Briefly, the target cells were labeled with chromium for 90 min. After washing three times, the cells were incubated with peptides at a concentration of 1 µg/ml for 90 min. The cells were washed again, counted, and then mixed with TIL586 or CTL clone 5 at the indicated ratio of effector : targets (E:T). Chromium release was measured after 4 h incubation. The peptides were synthesized by a solid-phase method using a peptide synthesizer (Model AMS 422, Gilson Co., Inc., Worthington, OH). Some peptides were purified by HPLC and had greater than 98% in purity. The mass of some peptides was confirmed by mass spectrometry analysis. For titration of the CAG-3 peptide recognized by TIL586, 586EBV B cells were incubated with various concentrations of the purified CAG-3 peptide or portion thereof. Percentage of specific lysis was determined from the equation (A-B)/(C-B) X 100 where A is lysis of 586EBV B cells by TIL586 or clone 5 in the presence of a peptide, B is spontaneous release from 586EBV B cells in presence of the same peptide but in the absence of effector cells, and C is the maximum chromium release. Cold target inhibition of cytolysis was performed using ⁵¹Cr-labeled 586mel or 624mel cells as "hot" targets and 586EBV B and T2 cells pulsed with peptides as "cold" targets.